ELECTRONIC MEETING
10-18 August 2022

## ESTIMATING POST-RELEASE MORTALITY OF LONG-LINE CAUGHT TROPICAL TUNAS IN THE PACIFIC OCEAN

WCPFC-SC18-2022/SA-IP-12

Jeff Muir ${ }^{1}$, Richard Barker ${ }^{2}$, Melanie Hutchinson ${ }^{3}$<br>Bruno Leroy ${ }^{4}$, Simon Nicol ${ }^{4}$, Joe Scutt Phillips ${ }^{4}$

[^0]
# Muir et al. (2022) Estimating post-release mortality of long-line caught tropical tunas in the Pacific Ocean. Fisheries Research, 249: 10619. [www.sciencedirect.com/science/article/abs/pii/S0165783621003222] 

## Estimating post-release mortality of long-line caught tropical tunas in the Pacific Ocean

Jeff Muir ${ }^{1,2 \dagger}$, Richard Barker ${ }^{3}$, Melanie Hutchinson ${ }^{4}$, Bruno Leroy ${ }^{1}$, Simon Nicol ${ }^{1,5}$, Joe Scutt Phillips ${ }^{1 \dagger^{*}}$<br>${ }^{1}$ The Pacific Community, BPD5 Noumea, New Caledonia<br>${ }^{2}$ 2651-C Waiomao Rd, Honolulu, HI 96816, USA<br>${ }^{3}$ Division of Sciences, University of Otago, Dunedin, New Zealand<br>${ }^{4}$ Cooperative Institute for Marine and Atmospheric Research, Pacific Islands Fisheries Science Center, 1845 Wasp Blvd. Bldg. 176 Honolulu, Hawaii 96818<br>${ }^{5}$ Centre for Conservation Ecology and Genomics, Institute for Applied Ecology, University of Canberra, Bruce 2617, Australia<br>${ }^{\dagger}$ These authors have contributed equally to this work and share first authorship<br>*Corresponding Author: joes@spc.int


#### Abstract

Post-release mortality experiments were undertaken on bigeye (Thunnus obesus) and yellowfin ( $T$. albacares) tuna to assess potential post-release survival probabilities of conventional tagging on such individuals during commercial longline operations. Survival was estimated from the release of 32 tuna tagged with pop-up, satellite survivorship tags (sPATs) during a commercial fishing trip aboard a Hawaii based tuna longline vessel. The tagged fish ranged in size from $103-145 \mathrm{~cm}$ fork length in size. Of the 32 sPATs released, 27 reported earlier than the pre-programmed 60 days, and five never reported. Of the 27 tags that reported data one tag appeared to shed early while the fish was still alive and swimming 32 days post-release while the remaining 26 tags deployed early due to mortality or other events, as observed through either sinking, ingestion by predators, or detached tags. Most mortalities occurred within four days post-release. The overall survival probability was estimated at around $10 \%$, with some evidence that reduced time on the hook may increase this probability up to $50 \%$. These results suggest that bigeye and yellowfin tuna caught during typical longline operations are unlikely to be suitable for electronic or conventional tagging experiments. Our study also suggests all discarded tuna should be treated as mortalities (including those discarded alive) for stock assessment purposes.


Key words: bigeye, yellowfin, sPAT, Hawaiian Islands, capture-mark-recapture

## 1. Introduction

Understanding post release mortality aids in the development of non-retention, catch and release policies and the design of monitoring programs for fisheries globally (Davis, 2002; Cooke and Shramm, 2007; Ellis et al., 2017; Reinhardt et al., 2017). Fishing mortality ( $F$ ) (of recovered gear) can be split into two components, at vessel ( $F_{\text {capture }}$ ) and, for animals released alive after capture, the mortality that may occur after release ( Freleases (Campana et al., 2009; Carruthers et al., 2009; Gilman et al., 2013; Musyl et al., 2011; 2015). $F_{\text {capture }}$ can be considered in its general form as either an instantaneous or
annual fishing mortality (Piet et al. 2007), typically estimated using the number of observed and recorded dead animals caught, whereas estimating $F_{\text {release }}$ is more challenging as individuals are no longer observed once they are released alive from the fishing gear. Estimated rates of $F_{\text {release }}$ vary considerably across taxa, gear types and environments (e.g. Eddy et al., 2016; Ellis et al., 2017; Musyl and Gilman, 2019; Nunes et al., 2019; Gil et al., 2020) and mortality due to the fishing interaction can be immediate or delayed for more than one-month (Swimmer et al., 2006; Musyl and Gilman, 2018; Schweitzer et al., 2020).

Non-retention of bycatch species often occurs in fisheries either for conservation purposes or the lack of a market for the product (Chan et al., 2014). 'High-grading' of target species can also occur for market purposes when the price for particular size classes are low in comparison to other sizes (Robertson, 1998; Huang and Liu, 2010). Estimates of non-retained catch and subsequent $F_{\text {release }}$ can be important for accurate stock and ecological risk assessments, particularly where non-retention rates are high (Cook, 2019) or in fisheries where quotas are derived from models that integrate total mortality information (Punt et al., 2006).

Frelease can also influence the design of Capture-Mark-Recapture (CMR) and biotelemetry studies (Brownie and Robson 1983). Such studies are widely used in fisheries science to estimate population parameters such as age/size specific mortalities (Polacheck et al., 2010), abundances (Schwarz and Seber, 1999; Bird et al., 2014), movement (Sibert and Hampton, 2003), and individual growth (Eveson et al., 2015). Increasingly CMR experiments are being applied in the research and management of tropical tuna fisheries (Fonteneau and Hallier, 2015; Leroy et al., 2015; Murua et al., 2015). The intent of CMR experiments is to mark a subset of the population and then monitor the population through recaptures of the marked fish. Population parameters can then be estimated for the size or age classes that the marked population aims to represent.

In the Western and Central Pacific Ocean (WCPO) the most applied method for capturing large numbers of live tuna for CMR has been the use of pole and line fishing techniques (Leroy et al., 2015). To a lesser extent, dangling, handline and sportfishing on targeted CMR trips for bigeye (Thunnus obesus) and yellowfin (T. albacares) has yielded some individuals in the 70-130 cm fork length (FL) in size, but not enough have been captured using these methods to form a meaningful conclusion on $F_{\text {release. }}$ Pole and line is the most effective fishing technique for fish less than 70 cm FL in size as they can be rapidly brought on board a fishing vessel in large numbers and in a condition suitable for marking (Leroy et al., 2015). For tropical tuna species such as bigeye and yellowfin tuna which can grow to lengths in excess of 150 cm FL, obtaining a tagged population that is representative of all age/size classes has been dependent on the marking of sufficient numbers of $<70 \mathrm{~cm}$ fish to permit a proportion of this marked cohort to survive until they reach larger size classes. Alternatively, these larger size classes can be caught, marked and released directly.

The largest commercial catches of bigeye and yellowfin tuna in the WCPO come from purse-seine and longline fisheries (Williams and Ruaia, 2020) with purse-seine vessels typically catching fish of <90 cm FL in length, and longline vessels catching those $>90 \mathrm{~cm}$ in length. Thus far, CMR experiments for yellowfin and bigeye in the WCPO have resulted in few recaptures of individuals in larger size classes (SPC-OFP, 2020). Consequently, assessments of these stocks have little CMR data to integrate for estimating mortalities associated with the longline fisheries. It remains unclear if the reason for low recaptures is due to an insufficient number of marked individuals of larger size classes being present in the population or that tag recaptures are simply under reported in longline fisheries. To address the former, longline caught individuals that are alive and considered in a healthy condition can be marked and released to increase tagged individuals in these size classes directly (ICCAT 2019).

In the WCPO in 2019 the non-retained catch of bigeye and yellowfin in longline fisheries was 2.4\% and $2.1 \%$ for each species respectively (Williams, 2020, Suppl. Mat. Figure S1). Although fisheries observers collect information on fish condition at release (alive healthy, alive injured and distressed, alive but dying, dead), all non-retained bigeye and yellowfin catches in the WCPO are assumed dead for stock assessment purposes (Williams, 2020).

Survivorship pop-up satellite tags have been used to estimate post-release mortality in tuna species caught within recreational fisheries (Kneebone et al. 2021). Here, we describe a similar study applied to longline caught and released bigeye and yellowfin tuna to investigate the $F_{\text {release }}$ of these species caught on longline fisheries operating in a commercial setting. We examine our results to inform (i) the design of future CMR experiments for bigeye and yellowfin tuna and (ii) total $F$ estimates used in stock assessments for these species.

## 2. Methods

### 2.1 Marking

Thirty-two survival pop-up archival transmitting tags (sPATs, Wildlife Computers, Inc. Redmond, WA) were deployed on bigeye and yellowfin tuna by a specialist tagging technician in the Hawaiian Islands Exclusive Economic Zone and neighbouring high seas between 18 November - 13 December 2020. The tags were deployed from the catch of a US-flagged fishing vessel during normal longline fishing activities.

The fishing vessel's longline consisted of 1000 lb test monofilament mainline, deployed and retrieved from a hydraulic spool (Lindgren Pitman (LP), Inc. Pompano Beach, FL), mounted on the aft portion of the house of vessel. The mainline was deployed on the port side of the vessel with a line shooter, also manufactured by LP. The speed of this line shooter can be adjusted so that gear can be targeted to different depths. The crew deployed plastic floats on 20 m float lines (per US regulations) and branch lines were 10 m long and composed of 2.1 mm diameter monofilament, to a 45-gram leaded swivel with a 30 cm braided wire trace and a baited 15/0 offset stainless steel ringed circle hook. Floats and branch lines were attached to the mainline at given time intervals, announced with a hook and float timer, and manufactured by LP. Additionally, five "radio" buoys were attached at the beginning and end of the line, and at each quarter. These buoys, manufactured by Marine Instruments, relay position, drift speed and direction, and water temperature every 10 minutes, and are displayed in a proprietary program on the vessel's bridge. This allowed the Captain to actively monitor the line's drift during and after the set and allow for gear recovery in the event of a parted mainline. Oceanographic factors, mainly sea-surface temperature, sea-surface height, and surface current were updated every second day on Orb-Image programming, and the captain identified favoured areas for setting considering these factors.

Setting occurred at or soon after daylight, depending on when the previous day's haul was completed. Haul completion was frequently late, causing the subsequent set to be delayed later in the day. Hauling on the starboard side of the vessel began at or around sunset and lasted 10-16 hrs. The vessel set 32-40 nm of horizontal distance (the actual distance of the mainline is much more due to the use of the line shooter) and 2500-3280 hooks which was typical of the Hawaii based longline fishery. Bait species used included pacific saury (Cololabis saira) and sardine (Sardinops sagax). Depth of set was targeted at or below the thermocline, which in most cases was 130-165 m, with 24 hooks per float.

Only fish considered as appropriate for tagging by the tagging technician were selected and tagged. During the hauling operation, tagging candidates were identified, if the fish, at leader was swimming and pulling strongly and did not have visible "line rash", and abrasion on the skin caused from the fish
being on the hook for an extended period of time and chafing against the line. Once the fish was identified as appropriate for tagging, a 120 cm half-moon shaped aluminium landing net with knotless webbing was used to bring the fish onboard (Figure 1). Because of the size of the tuna being landed (some in excess of 65 kg ), at least 3 crewmen were required to lift the netted fish through the door and carefully place it on the matted deck for tagging. After another quick assessment for any other obvious damage to the fish (eye damage, excessive bleeding), the hook was removed, and the sPAT was placed in the pterigiophores of the second dorsal fin using a titanium pronged applicator. On the opposite side of the second dorsal fin, a 13 cm yellow conventional dart tag with recapture information was also placed. The fish was measured to the nearest centimetre using an aluminium calliper and then gently released overboard, with time and apparent condition at release noted. Species, float number, hook number and time and date of release were also recorded. The sPATs were programmed to "pop-off" 60 days after deployment. Each sPAT was rigged using a 10 cm monofilament tether and titanium anchor.

### 2.2 Survival PAT Data

The 32 sPATs were programmed to burn through their tether pin to "pop-up" on either the $60^{\text {th }}$ day after deployment, once a critical depth limit of 1700 m was reached, or once the tag was floating at the surface for 24 hours. On reporting, sPATs provided daily minimum and maximum depths and external temperatures, alongside the relative change in ambient light level during the entire deployment. Higher resolution depth and temperature data from the final 5 days prior to reporting were also transmitted to aid in classification of fate and behaviour.

The data from each reporting sPAT was examined to determine the fate of the tagged fish. Fate was classified as death through either predation or stress/injury following the longline capture and tagging process, or survival through the SPAT either reporting at the programmed 60 -days of liberty or with evidence of erroneous, early detachment. Predation was identified by ingestion of the tag, indicated by reduced changes in ambient light and temperature, followed by expulsion and floating of the tag to enable its reporting via satellite. When fate was classified as death, the number of days of survival was estimated to the nearest day by examination of the transmitted behavioural data.

### 2.3 Data Analyses

Cox regression models were fitted to the data in a survival analysis, using the number of days until apparent mortality as the response variable, and with tags that appeared to indicate survival to the end of the 60 -day reporting period treated as right-censored. Species, length, and recorded fish condition were all considered as potential predictors of death rate. The effect of float position in the set at which the fish was landed, and hook number from float were also examined, as a potential proxy for time spent on the hook and depth, respectively.

## 3. Results

## 3.1 sPAT Reporting

During the 25 -day cruise, 15 normal length sets and one short set (Suppl. Mat. Table S1) were made and nearly 6000 kgs of pelagic fish were landed. In addition to the primary target of bigeye tuna, 12 other marketable species of pelagic fish, three unmarketable, and five species of pelagic sharks were caught during the trip. sPATs were deployed on 26 bigeye and six yellowfin tuna (Suppl. Mat. Table S2).

Of the 32 sPAT tags released, 27 reported earlier than the pre-programmed 60 days, and five never reported. The five non-reporting tags may have been failures, although on examination of the depth
profiles for those fish that died and sank following longline capture and tagging, the sinking speed was fast, often in excess of $0.3 \mathrm{~m} / \mathrm{s}$. The sPAT tags used in this study are tested to survive pressures of 2000-2500m but, given the observed sinking speed and pin burn times in reporting tags, it is possible that the sPATs that did not report were crushed at depth before they were able to release and float to the surface. In our subsequent analyses, these five non-reporting tags can be treated as censored survivors at time zero, effectively removing them from the analysis as they contribute no information on post-release mortality rates.

Of the sPAT tags that reported, all but one indicated detachment through either sinking or predation, the majority of which occurred within four days (Figure 2). The remaining sPAT that reported appeared to indicate a pre-mature release where the tag was shed 32 days after tagging (fish 208712, Suppl. Mat. Figure S2). The longest survival time was 49 days from release, for a bigeye tuna that was exhibiting typical, and diurnal behaviour (Scutt Phillips et al. 2017) immediately prior to apparently dying at depth and sinking to the ocean floor (fish 208733, Suppl. Mat. Figure S3). A summary of release covariates and survival times is given in Table 1.

Detailed examination of the vertical behaviour provided by reporting sPATs revealed three main mortality patterns (Figure 3). The most common was an extended surface-association up to three- or four-days post release before apparent mortality and sinking to the ocean floor ( 12 fish). There were also several clear predation events within the few days post-release, indicated by tags reporting a lack of changing light levels, some tags reporting consistent temperatures with depth, and extended, nontuna like vertical movement behaviour (six fish). These included very shallow vertical behaviour with short, bounce dives (similar to that reported for billfish, e.g. Sippel et al., 2011, Luo et al., 2006, Hoolihan et al., 2011, or possible beaked whales e.g. Baird et al. 2006), and high frequency vertical movement with a diel pattern similar to that reported for silky shark (Carcharhinus falciformis) behaviour, (Filmalter et al., 2011; 2015), and shortfin mako (Isurus oxyrinchus) shark (Abascal et al., 2011). Finally, there were four instances of atypical behaviour in the days immediately following release, ending with sudden floating of the tag at surface and one or more spike dives to depth before the tag began to report. While it is possible that these are shedding events, without considerable necrosis around the tagging area, we suggest that shedding of titanium anchors from behind the pterygiophores was less likely, given that the tagging technician did not report poor placement for these tags. Instead, we have assumed for the purposes of our analyses, that these are either predation events in which the tag is not ingested, or that the tag itself is predated upon and torn from the fish, before subsequently floating to the surface and then pulled down to depth or swallowed and immediately regurgitated by subsequent scavengers.

### 3.2 Post-release Mortality Estimates

As a supplement to Cox-regression modelling, the Kaplan-Meier estimator (reference) was used to provide a simple description of post-release mortality using data from 27 of the sPAT deployments. For this analysis the five non-reporting tags were excluded from the analysis. Given the uncertainty in assigning fates to four of the prematurely floating tags described above, two datasets were considered for analysis. A conservative dataset, assuming 26 observed mortality events, including these premature floating tags, and one censored event at day 32 corresponding to the apparent shedding event. An alternative survival analysis was also run on a less-conservative dataset, where the four prematurely floating tags were assumed to be shedding, and therefor censored, events and not predation mortalities.

Figure 4 shows the Kaplan-Meier survival curve for days since release for the conservative dataset. The observed survival status at the end of the 60-day study period was zero for all fish. Given the low
sample number in our analysis, identifying an accurate estimate of survival probability was not possible, but the data indicate an asymptote at 0.1-0.15. The median survival time, the point at which survival probability fell below 0.5 , occurred at two days, occurring between one and three days within $95 \%$ confidence limits.

When a dataset assuming floating tags as censored events rather than mortalities was used, the survival asymptote increased to around 0.2. Median survival time remained at two days, although the $95 \%$ confidence limits extended from one to six days.

### 3.3 Factors influencing mortality

Using Cox proportional hazards methods, four covariates were examined for their potential influence on survival in the most conservative dataset: species, fork length, condition at release, and float position on the line during hauling. However, given the number of observed mortalities in this experiment, there was little signal for a Cox regression survival analysis to identify significant effects.

Testing of the proportional hazards assumptions and other model diagnostics were also extremely limited for the same reasons. While many of the fish in this study were tagged during the same haul event, and then released into potentially similar environments of predation, we assume here that all survival times are independent. Schoenfield residuals (Grambsch \& Therneau 1994) were calculated for each covariate to examine the hazard response over time (see Supplementary materials figure S4). Fish condition was the only covariate that indicated breaking of the proportional hazards assumptions ( $\mathrm{p}<0.01$ ), although this appeared driven by the very low numbers of events occurring after the initial four days.

Regardless, none of the covariates were found to be significant in estimated survival probability when using the conservative dataset. However, when considering early-release floating tags as censored events, float number was found to be significant (hazard ratio $=1.012,1.001$ to $1.02495 \% \mathrm{Cl}, \mathrm{P}=0.04$ ), with decreased survival predicted when tagged fish were hauled later in the set. Examining model predicted survival curves across the quantile values of the observed float number in our study, survival probability ranged from around 0.5 for fish tagged at the beginning of line hauling, to 0.05 for fish hauled later in the set (figure 5).

## 4. Discussion

Our study indicates that post release mortality of tropical tuna caught by commercial longline gear and operations is likely to be very high. The survival analyses indicate a $20-30 \%$ survival rate within four days of release, potentially reaching an asymptote of just $10 \%$. Survival rates of large tunas satellite tagged using rod and reel gears have been estimated as much higher than those found in longline captured tuna here ( $92 \%$, Kneebone et al. 2021). Other studies of tagged tuna captured by rod and reel also indicate that tagging itself does not result in high levels of mortality (e.g. Lam et al. 2020 Tolotti et al. 2020), and lab studies also showed no survival or behavioural impacts of satellite tagged, captive mahimahi (Coryphaena hippurus) compared to untagged conspecifics (McGuigan et al. 2021). It appears that the commercial longline hauling is more responsible for the high mortality we have observed, rather than the satellite tagging procedure itself. These results suggest that targeting bigeye and yellowfin tuna for mark recapture through standard commercial longlining operations is not a viable method of obtaining growth or other life history information of these size classes, regardless of the type of tag deployed. Despite individuals being selected by scientific crew as in the best possible condition for tagging, examination of the vertical behaviour reported by sPATs post-release suggest that the probability of post-release survival was extremely low. Either the time on the hook, hauling and capture simply left fish too exhausted or stressed to survive release back
into their environment (Pankhurst \& Dedual 1994), or there was a high incidence of serious injuries that were not apparent from an external examination. Individuals either struggled with depth-control before sinking or were quickly predated by what appears to be sharks or marine mammals, which may have been present in high densities around the hauling operation of the fishing vessel. For either case, this happened within four days for $80 \%$ of fish. After that we observed three instances of potential, 'delayed' mortality ranging from 6 to 26 days, and one clear instance of predation just prior to day five. Natural mortality for bigeye and yellowfin tuna at these size classes is estimated to be around $3.5-4 \%$ and $6-10 \%$, respectively, per month (Decharme-Barth et al., 2020; Vincent et al., 2020), and it is worth noting that one of the two fish tagged in this study assumed not to have experienced longlinetagging mortality still appeared to die of unassigned causes nearly two months after release. It is plausible that this represented a natural mortality event.

High rates of post release mortality have not been observed in other species caught by pelagic longline fisheries. Pelagic sharks are reported to experience post release rates ranging from $11 \%$ to $57 \%$ out to 30 days depending on species, animal condition, handling, and fishery (Musyl and Gilman, 2019; Hutchinson et al., 2021) with slightly lower rates reported for billfishes (11\%-18\%) (Musyl and Gilman, 2019). These rates are approximately equivalent to those reported in recreational fisheries for tuna and billfishes (Sepulveda et al. 2020; Tracey et al., 2016; Marcek and Graves, 2014; Stokesbury et al., 2011; Horodysky and Graves 2005; Musyl et al., 2015; Domeier et al., 2003). Hook types can significantly affect the mortality rates of species caught by longline fisheries (Reinhardt et al., 2018). For example, lower rates of $F_{\text {capture }}$ have been observed for yellowfin and bluefin tuna when circlehooks in comparison to J-hooks are deployed (Reinhardt et al., 2018) indicating that these hook types may have lower impact on these species. In the same meta-analyses however, Reinhardt et al., (2018) were not able to detect an effect of hook type on bigeye tuna. The technique used to lift fish from the water to the deck can also influence rates of post release mortality (Sakai and Itoh, 2013), and it is very possible the hauling and de-hooking procedure specific to our tagging study also influenced postrelease mortality. Mortalities were higher for southern bluefin tuna when lifted using the branch line in comparison to the lift occurring via a spoon-net or basket (Sakai and Itoh, 2013), and the half-moon shaped landing net used in our study may have similarly aided in reducing the stress associated with lifting on to the deck for tagging. However, while other potential covariates were collected in our study, including those that may be proxy for time spent on the hook by individual fish, our results did not show strong evidence for an effect on mortality when compared to the null model of survival and many more replicates would be required to quantify this further. Such an experiment appears to be of little worth, given the very low survival rates we have presented here.

Our analysis supports the current approach of assuming that all longline captures of tuna (i.e. including live discards) as mortalities for the purposes of constructing catch data for stock assessments of tunas in the WCPO (Williams, 2020). Increasing access to tuna has been identified as a potential solution to improve the food security within the Pacific Islands and nutrition of its communities (Bell et al., 2015). While discarding of tuna in the WCPO more commonly occurs on catches of small-sized tuna (Chan et al., 2014) some discarding of large sized fish occurs in the longline fisheries (Williams, 2020, Suppl. Mat. Figure S1). Given the high mortalities likely experienced, a full retention (Chan et al., 2014) or other policy that incentivizes their use in supplementing Pacific Island food security needs may be a better use of this resource.

Conventional tagging of tropical tunas caught on longline gears is a technique employed by some tagging programmes (e.g. the Atlantic Ocean Tuna Tagging Programme), although at a small scale (< 500 releases, AOTTP data https://www.iccat.int/aottp/en/aottp-data-release.html). Short lines or short surface longlines, less than 500 hooks (Itano, 2004) has also successfully been used to capture,
tag and release tunas where post release mortalities have been low (Allain et al., 2005; Williams et al., 2014), but these trips have been dedicated tagging expeditions using modified techniques rather than commercial operations. While the longline gear settings varied between the different platforms used in these expeditions, the operations were specifically adapted to optimize tagged fish survival. A low number of hooks (175 to 270) per set, and caution taken in the hauling speed to minimize fish barotrauma was conducted (Allain et al., 2005). The significance of float number in our analysis of the less conservative dataset presented here suggests that such short sets and less hauling stress may indeed decrease post-release mortality.

While modified fishing operations may approach results in lower rates of $F_{\text {capture }}$ and $F_{\text {release }}$ in comparison to standard commercial operations, the small number of fish captured with these modifications is likely insufficient for broad scale application of conventional tagging. Our results highlight the importance of large-scale CMR of small individuals to ensure significant numbers of fish surviving to adult life stages, coupled with active engagement in the recovery of tags in longline fisheries.

## Acknowledgements

The authors thank the assistance of the captain and crew of the fishing vessel to implement the tagging of the tuna for this study. NOAA Observer, Ms Ashley Graham provided assistance with data collection on board the vessel. Funding was provided by the Western and Central Pacific Fisheries Commission and the European Union "Pacific-European-Union-Marine-Partnership" programme.

JSP conceived the project, JSP, SN, BML, JM designed the project, JM implemented the project, RB advised on analyses, MH facilitated access to observer data and provided equipment. All authors contributed to writing and editing the manuscript.

## References

Abascal, F.J., Quintans, M., Ramos-Cartelle, A., Mejuto, J., 2011. Movements and environmental preferences of the shortfin mako, Isurus oxyrinchus, in the southeastern Pacific Ocean. Mar. Biol. 158, 1175-1184. DOI 10.1007/s00227-011-1639-1.

Allain, G., Lehodey, P., Kirby, D.S., Leroy, B., 2005. The influence of the environment on horizontal and vertical bigeye tuna movements investigated by analysis of archival tag records and ecosystem model outputs. WCPFC-SC01-BI-WP-03. $1^{\text {st }}$ Regular Session of the Scientific Committee of the Western and Central Pacific Fisheries Commission, Noumea, New Caledonia, 8-19 August 2005.

Baird, R.W., Webster, D.L., McSweeney, D.J., Ligon, A.D., Schorr, G.S. and Barlow, J., 2006. Diving behaviour of Cuvier's (Ziphius cavirostris) and Blainville's (Mesoplodon densirostris) beaked whales in Hawai'i. Canadian Journal of Zoology, 84(8), pp.1120-1128.

Bell, J.D., Allain, V., Allison, E.H., Andréfouët, S., Andrew, N.L., Batty, M.J., Blanc, M., Dambacher, J.M., Hampton, J., Hanich, Q., Harley, S., Lorrain, A., McCoy, M., McTurk, N., Nicol, S., Pilling, G., Point, D., Sharp, M.K., Vivili, P., Williams, P., 2015. Diversifying the use of tuna to improve public health in Pacific Island countries and territories. Mar. Pol. 51, 584-591.
http://dx.doi.org/10.1016/j.marpol.2014.10.005.
Bird, T., Lyon, J., Nicol, S., McCarthy, M., Barker, R., 2014. Estimating population size in the presence of temporary migration using a joint analysis of telemetry and capture-recapture data. Methods Ecol. Evol. 5, 615-625. https://doi.org/10.1111/2041-210X.12202.

Brownie, C., Robson, D. S., 1983. Estimation of time—specific survival rates from tag-resighting samples: a generalization of the Jolly-Seber model. Biometrics 39, 437-453.

Campana, S.E., Joyce, W., Manning, M.J., 2009. Bycatch and discard mortality in commercially caught blue sharks Prionace glauca assessed using archival satellite pop-up tags. Mar. Ecol. Prog. Ser. 387, 241-253. DOI: https://doi.org/10.3354/meps08109.

Carruthers, E.H., Schneider, D.C., Neilson, J.D., 2009. Estimating the odds of survival and identifying mitigation opportunities for common by catch in pelagic longline fisheries. Biol. Conserv. 142, 26202630. https://doi.org/10.1016/j.biocon.2009.06.010.

Chan, V., Clarke, R., Squires, D., 2014. Full retention in tuna fisheries: benefits, costs and unintended consequences. Mar. Pol. 45, 213-221. https://doi.org/10/1016/j.marpol.2013.10.016.

Cook, R.M., 2019. Inclusion of discards in stock assessment models. Fish Fish. 20, 1232-1245. https://doi.org/10.1111/faf. 12408.

Cooke, S.J., Shramm, H.L., 2007. Catch-and-release science and its application to conservation and management of recreational fisheries. Fish. Manag. Ecol. 14, 73-79. https://doi.org/10.1111/j.13652400.2007.00527.x.

Davis, M.W., 2002. Key principles for understanding fish bycatch discard mortality. Can. J. Fish. Aquat. Sci. 59, 1834-1843. DOI:10.1139/F02-139.

Ducharme-Barth, N., Vincent, M., Hampton, J., Hamer, P., Williams, P. and Pilling, G., 2020. Stock assessment of bigeye tuna in the western and central Pacific Ocean. WCPFC-SC16-2020/SAWP-03 (Rev. 01), 144.

Domeier, M.L., Dewar, H., and Nasby-Lucas, N., 2003. Mortality rate of striped marlin (Tetrapturus audax) caught with recreational tackle. Mar. Freshw. Res. 54, 435-445. https://doi.org/10.1071/MF01270.

Eddy, C., Brill, R., Bernal, D., 2016. Rates of at-vessel mortality and post-release survival of pelagic sharks captured with tuna purse seines around drifting fish aggregating devices (FADs) in the equatorial eastern Pacific Ocean. Fish. Res. 174, 109-117. 10.1016/j.fishres.2015.09.008.

Ellis, J.R., Mccully Phillips, S.R., Poisson, F., 2017. A review of capture and post-release mortality of elasmobranchs. J. Fish. Biol. 90, 653-722. http://dx.doi.org/10.1111/jfb.13197.

Eveson, J.P., Million, J., Sardenne, F., Le Croizier, G., 2015. Estimating growth of tropical tunas in the Indian Ocean using tag-recapture data and otolith-based age estimates. Fish. Res. 163, 58-68. https://doi.org/10.1016/j.fishres.2014.05.016.

Filmalter, J.D., Dagorn, L., Cowley, P.D., Taquet, M., 2011. First descriptions of the behavior of silky sharks, Carcharhinus falciformis, around drifting fish aggregating devices in the Indian Ocean. Bull. Mar. Sci. 87, 325-337. https://doi.org/10.5343/bms.2010.1057.

Filmalter, J., Cowley, P., Forget, F., Dagorn, L., 2015. Fine-scale 3-dimensional movement behaviour of silky sharks Carcharhinus falciformis associated with fish aggregating devices (FADs). Mar. Ecol. Prog. Ser. 539, 207-223. DOI: https://doi.org/10.3354/meps11514.

Fonteneau, A., Hallier, J-P., 2015. Fifty years of dart tag recoveries for tropical tuna: A global comparison of results for the western Pacific, eastern Pacific, Atlantic, and Indian Oceans. Fish. Res. 163, 7-22. https://doi.org/10.1016/j.fishres.2014.03.022.

Gil, M.d.M., Palmer, M., Morey G., Manjabacas A., Pastor E., Díaz-Gil C., Grau, A.M., 2020. Assessing relative post-release mortality for the transparent goby fishery: Environmental drivers and the utility of vitality metrics. PLoS ONE 15, e0230357. https://doi.org/10.1371/journal.pone.0230357.

Gilman, E., Suuronen, P., Hall, M., Kennelly, S., 2013. Causes and methods to estimate cryptic sources of fishing mortality. J. Fish. Biol. 83, 766-803. https://doi.org/10.1111/jfb.12148.

Grambsch, P. M., \& Therneau, T. M. (1994). Proportional hazards tests and diagnostics based on weighted residuals. Biometrika, 81(3), 515-526.

Hoolihan, J.P., Luo, J., Goodyear, C.P., Orbesen, E.S., Prince, E.D., 2011. Vertical habitat use of sailfish (Istiophorus platypterus) in the Atlantic and eastern Pacific, derived from pop-up satellite archival tag data. Fish. Oceanogr. 20, 192-205. https://doi.org/10.1111/j.1365-2419.2011.00577.x.

Horodysky, A.Z., and Graves, J.E., 2005. Application of pop-up satellite tag technology to estimate postrelease survival of white marlin (Tetrapterus albidus) caught on circle and straight-shank "J" hooks in the western North Atlantic recreational fishery. Fish. Bull. 103, 84-96.

Huang, H-W., and Liu, K-M., 2010. Bycatch and discards by Taiwanese large-scale tuna longline fleets in the Indian Ocean. Fish. Res. 106, 261-270. DOI:10.1016/j.fishres.2010.08.005.

Hutchinson, M., Siders, Z., Stahl, J., Bigelow, K., 2021. Quantitative estimates of post-release survival rates of sharks captured in Pacific tuna longline fisheries reveal handling and discard practices that improve survivorship. PIFSC Data Report DR-21-001. https://doi.org/10.25923/0m3c-2577.

ICCAT (International Commission for the Conservation of Atlantic Tunas), 2019. AOTTP Newsletter 0109, https://issuu.com/aottp.iccat/docs/newsletter 01-09.

Itano, D., 2014. Hawaiian-style small-scale deep setting longline technique used on seamounts. SPC Fisheries Newsletter 111, 21-26.

Kneebone, J., Benoît, H.P., Bernal, D. and Golet, W., 2021. Application of a parametric survival model to understand capture-related mortality and predation of yellowfin tuna (Thunnus albacares) released in a recreational fishery. Canadian Journal of Fisheries and Aquatic Sciences, 78(4), pp.386-399.

Lam, C.H., Tam, C., Kobayashi, D.R. and Lutcavage, M.E., 2020. Complex Dispersal of Adult Yellowfin Tuna From the Main Hawaiian Islands. Frontiers in Marine Science, 7, p.138.

Leroy, B., Nicol, S., Lewis, A., Hampton, J., Kolody, D., Caillot, S., Hoyle, S., 2015. Lessons learned from implementing three, large-scale tuna tagging programmes in the western and central Pacific Ocean. Fish. Res. 163, 23-33. https://doi.org/10.1016/j.fishres.2013.09.001.

Luo, J., Prince, E.D., Goodyear, C.P., Luckhurst, B.E., Serafy, J.E., 2006. Vertical habitat utilization by large pelagic animals: a quantitative framework and numerical method for use with pop-up satellite tag data. Fish. Oceanogr. 15, 208-229. doi:10.1111/j.1365-2419.2006.00360.x.

Marcek, B.J., and Graves, J.E., 2014. An estimate of postrelease mortality of school-size bluefin tuna in the U.S. recreational troll fishery. N. Am. J. Fish Manage. 34, 602-608. DOI: 10.1080/02755947.2014.902411.

McGuigan, C.J., Schlenker, L.S., Stieglitz, J.D., Benetti, D.D. and Grosell, M., 2021. Quantifying the effects of pop-up satellite archival tags on the swimming performance and behavior of young-adult mahi-mahi (Coryphaena hippurus). Canadian Journal of Fisheries and Aquatic Sciences, 78(1), pp.3239.

Murua, H., Eveson, J.P., Marsac, F. 2015. The Indian Ocean Tuna Tagging Programme: Building better science for more sustainability. Fish. Res 163, 1-6. DOI:10.1016/j.fishres.2014.07.001

Musyl, M.K., Brill, R.W., Curran, D.S., Fragoso, N.M., McNaughton, L.M., Neilsen, A., Kikkawa, B.S., Moyes, C.D., 2011. Postrelease survival, vertical and horizontal movements, and thermal habitats of five species of pelagic sharks in the central Pacific Ocean. Fish. Bull. 109, 341-368.

Musyl, M.K., Moyes, C.D., Brill, R.W., Mourato, B.L., West, A., McNaughton, L.M., Chiang, W-C., Sun, C-L., 2015. Postrelease mortality in istiophorid billfish. Can. J. Fish. Aquat. Sci. 72, 538-556. https://doi.org/10.1139/cjfas-2014-0323.

Musyl, M.K., Gilman, E.L., 2018. Post-release fishing mortality of blue (Prionace glauca) and silky shark (Carcharhinus falciformes) from a Palauan-based commercial longline fishery. Rev. Fish. Biol. Fisheries 28, 567-586. https://doi.org/10.1007/s11160-018-9517-2.

Musyl, M.K., Gilman, E.L., 2019. Meta-analysis of post-release fishing mortality in apex predatory pelagic sharks and white marlin. Fish Fish. 20, 466-500. https://doi.org/10.1111/faf. 12358.

Nunes, D.M., Hazin, F.H.V., Branco-Nunes, I.S.L., Hazin, H., Pacheco, J.C., Afonso, A.S., Mourato, B.L., Carvalho, F.C., 2019. Survivorship of species caught in a longline tuna fishery in the western equatorial Atlantic Ocean. Lat. Am. J. Aquat. Res. 47, 798-807. http://dx.doi.org/10.3856/vol47-issue5-fulltext-9.

Pankhurst, N.W. and Dedualj, M., 1994. Effects of capture and recovery on plasma levels of cortisol, lactate and gonadal steroids in a natural population of rainbow trout. Journal of Fish Biology, 45(6), pp.1013-1025.

Polacheck, T., Eveson, J.P., Laslett, G.M., 2010. Classifying tagging experiments for commercial fisheries into three fundamental types based on design, data requirements and estimable population parameters. Fish Fish. 11, 133 - 148. DOI:10.1111/j.1467-2979.2010.00360.x.

Punt, A.E., Smith, D.C., Tuck, G.N., Methot, R.D., 2006. Including discard data in fisheries stock assessments: two case studies from south-eastern Australia. Fish. Res. 79, 239-250. https://doi.org/10.1016/j.fishres.2006.04.007.

Reinhardt, J.F., Weaver, J., Latham, P.J., Andrea Dell'Apa, A., Serafy, J.E., Browder, J.A., Christman, M., Foster, D.G., Blankinship, D.R., 2017. Catch rate and at-vessel mortality of circle hooks versus J-hooks in pelagic longline fisheries: A global meta-analysis. Fish Fish. 2017, 1-18. DOI: 10.1111/faf. 12260.

Robertson, G., 1998. The culture and practice of longline tuna fishing: implications for seabird by-catch mitigation. Bird Conserv. Int. 8, 211-221. https://doi.org/10.1017/S095927090000188X.

Sakai, O. Itoh, T., 2013. Post-releases survival of Southern Bluefin Tuna released from longline vessels. CCSBT-ESC/1309/34.

Schweitzer, C.C., Horodysky, A.Z., Price, A.L., Stevens, B.G., 2020. Impairment indicators for predicting delayed mortality in black sea bass (Centropristis striata) discards within the commercial trap fishery. Conserv. Physiol. 8, 2051-1434. https://doi.org/10.1093/conphys/coaa068.

Schwarz, C., Seber, G.A.F., 1999. Estimating Animal Abundance: Review III. Stat. Sci. 14, 427-456. https://www.jstor.org/stable/2676809.

Scutt Phillips, J., Pilling, G.M., Leroy, B., Evans, K., Usu, T., Lam, C.H., Schaefer, K.M. and Nicol, S., 2017. Revisiting the vulnerability of juvenile bigeye (Thunnus obesus) and yellowfin (T. albacares) tuna
caught by purse-seine fisheries while associating with surface waters and floating objects. PloS one, 12(6), p.e0179045.

Sepulveda, C. A., Aalbers, S. A., Wang, M., Kneebone, J., \& Bernal, D. 2020. Post-release survivorship of Pacific bluefin tuna (Thunnus orientalis) captured in the California recreational fishery. Fisheries Research, 223, 105413.

Sibert, J., Hampton, J., 2003. Mobility of tropical tunas and the implications for fisheries management. Mar. Pol. 27, 87-95. https://doi.org/10.1016/S0308-597X(02)00057-X.

Sippel, T., Holdsworth, J., Dennis, T., Montgomery, J., 2011. Investigating behaviour and population dynamics of striped marlin (Kajikia audax) from the southwest Pacific Ocean with satellite tags. PLoS One 6, e21087. https://doi.org/10.1371/journal.pone.0021087.

SPC-OFP, 2020. Project 42: Pacific Tuna Tagging Project Report and Work Plan for 2020-2023. WCPFC-SC16-RP-PTTP-01. 16th Regular Session of the Scientific Committee of the Western and Central Pacific Fisheries Commission, Pohnpei, Federated States of Micronesia, 12-19 August 2020.

Stokesbury, M.J.W., Neilson, J.D., Susko, E., and Cooke, S.J., 2011. Estimating mortality of Atlantic bluefin tuna (Thunnus thynnus) in an experimental recreational catch-and-release fishery. Biol. Conserv. 144, 2684-2691. https://doi.org/10.1016/j.biocon.2011.07.029.

Swimmer, Y., Arauz, R., McCracken, M., McNaughton, L., Ballestero, J., Musyl, M., Bigelow, K., Brill, R., 2006. Diving behavior and delayed mortality of olive ridley sea turtles Lepidochelys olivacea after their release from longline fishing gear. Mar. Ecol. Prog. Ser. 323, 253-261. doi:10.3354/meps323253.

Tolotti, M.T., Forget, F., Capello, M., Filmalter, J.D., Hutchinson, M., Itano, D., Holland, K. and Dagorn, L., 2020. Association dynamics of tuna and purse seine bycatch species with drifting fish aggregating devices (FADs) in the tropical eastern Atlantic Ocean. Fisheries Research, 226, p. 105521.

Tracey, S., Hartmann, K., Leef, M., McAllister, J., 2016. Capture-induced physiological stress and postrelease mortality for Southern bluefin tuna (Thunnus maccoyii) from a recreational fishery. Can. J. Fish. Aquat. Sci. 73, 1547-1556. https://doi.org/10.1139/cjfas-2015-0516.

Vincent, M., Ducharme-Barth, N., Hamer, P., Hampton, J., Williams, P. and Pilling, G., 2020. Stock assessment of yellowfin tuna in the western and central Pacific Ocean. WCPFC-SC16-2020/SAWP-04 (Rev. 01), 151.

Williams, P., 2020. Estimates of annual catches in the WCPFC statistical area. WCPFC-SC16-ST-IP-01. 16th Regular Session of the Scientific Committee of the Western and Central Pacific Fisheries Commission, Pohnpei, Federated States of Micronesia, 12-19 August 2020.

Williams, P., Ruaia T., 2020. Overview of tuna fisheries in the Western and Central Pacific Ocean, including economic conditions - 2019 (24July2020) - Rev.03. WCPFC-SC16-GN-IP-01. 16th Regular Session of the Scientific Committee of the Western and Central Pacific Fisheries Commission, Pohnpei, Federated States of Micronesia, 12-19 August 2020.


Figure 1. Half moon aluminium landing net used to bring yellowfin and bigeye tuna deemed suitable for tagging aboard.


Figure 2. Distribution of survival time for all deployed and reported sPATs for adult bigeye and yellowfin tuna captured by longline.

Table 1. Summary of release, fate, and covariates of tagged longline caught fish

| PTT <br> Tag No. | Species | Fork Length | $\begin{aligned} & \hline \text { Date } \\ & 2020 \end{aligned}$ | Fish Condition | Survived | Mortality due to Capture \& Tagging | Days <br> Survived | Float <br> No. | Hook <br> No. |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 170602 | Bigeye | 119 | 9/12 | Bleeding | 0 |  | 3 | 40 | 9 |
| 170603 | Bigeye | 118 | 9/12 |  | 0 |  | 0 | 48 | 6 |
| 208704 | Bigeye | 103 | 25/11 | Eye damage | 0 |  | 2 | 66 | 14 |
| 208705 | Bigeye | 127 | 29/11 |  | 0 |  | Unknown | 101 | NA |
| 208706 | Yellowfin | 117 | 22/11 |  | 0 |  | 18 | 18 | 17 |
| 208707 | Bigeye | 127 | 28/11 |  | 0 |  | Unknown | 60 | 14 |
| 208708 | Bigeye | 124 | 25/11 |  | 0 |  | 2 | 49 | 4 |
| 208709 | Bigeye | 127 | 6/12 |  | 0 |  | Unknown | 55 | 15 |
| 208710 | Bigeye | 145 | 5/12 |  | 0 |  | 2 | 93 | 16 |
| 208711 | Bigeye | 127 | 7/12 | Slow on release | 0 |  | 1 | 55 | 16 |
| 208712 | Bigeye | 113 | 27/11 | Slow on release | 1 |  | Unknown | 19 | 6 |
| 208713 | Bigeye | 127 | 4/12 | Slow on release | 0 |  | 0 | 89 | 23 |
| 208714 | Bigeye | 126 | 7/12 | Bleeding | 0 |  | 2 | 72 | 6 |
| 208715 | Bigeye | 132 | 30/11 |  | 0 |  | 3 | 2 | 12 |
| 208716 | Yellowfin | 136 | 7/12 |  | 0 |  | 0 | 74 | 23 |
| 208717 | Yellowfin | 116 | 25/11 |  | 0 |  | 6 | 58 | 23 |
| 208718 | Bigeye | 114 | 3/12 | Eye damage | 0 |  | 0 | 78 | 11 |
| 208719 | Yellowfin | 124 | 26/11 | Slow on release | 0 |  | 0 | 45 | 21 |
| 208720 | Bigeye | 120 | 27/11 | Slow on release | 0 |  | 26 | 116 | 9 |
| 208721 | Bigeye | 129 | 29/11 |  | 0 |  | 4 | 71 | 6 |
| 208722 | Bigeye | 130 | 4/12 |  | 0 |  | 1 | 80 | 12 |
| 208723 | Bigeye | 126 | 29/10 |  | 0 |  | 2 | 110 | 9 |
| 208724 | Yellowfin | 134 | 30/11 | Slow on release | 0 |  | Unknown | 71 | 11 |
| 208725 | Bigeye | 127 | 3/12 |  | 0 |  | 0 | 86 | 9 |
| 208726 | Bigeye | 126 | 7/12 | Slow on release | 0 |  | 1 | 92 | 17 |
| 208727 | Bigeye | 119 | 3/12 |  | 0 |  | 1 | 80 | 16 |
| 208728 | Bigeye | 124 | 30/11 |  | 0 |  | 4 | 5 | 14 |
| 208729 | Bigeye | 117 | 1/12 | Slow on release | 0 |  | 1 | 73 | 16 |
| 208730 | Bigeye | 103 | 22/11 |  | 0 |  | 1 | 31 | 15 |
| 208731 | Bigeye | 139 | 2/12 | Slow on release | 0 |  | Unknown | 32 | NA |
| 208732 | Yellowfin | 136 | 3/12 | Slow on release | 0 |  | 0 | 15 | 16 |
| 208733 | Bigeye | 120 | 29/11 | Slow on release | 0 |  | 49 | 5 | 9 |



Figure 3. Examples of classified bigeye tuna mortalities from pre-pop up, high resolution dive profiles prior to sPAT data transmission. Clockwise from top left: Depth holding with extended shallow behaviour prior to sinking, ingestion by a surface oriented, shallow-swimming predator, ingestion (inferred by light level data) by a predator with diurnal behaviour, premature release and floating of the tag followed by something dragging the tag to depth.


Figure 4. Kaplan-Meier survival curve using a conservative dataset of all reporting sPAT tags for adult bigeye and yellowfin tuna captured by longline.


Figure 5. Kaplan-Meier estimated survival curves when assuming premature floating tags as censored events and float number as a covariate of the hazard function. Estimated survival is shown with increasing float number (later haul time).


[^0]:    ${ }^{1}$ Independent fisheries consultant for the Pacific Community (SPC), Oceanic Fisheries Programme
    ${ }^{2}$ Division of Sciences, University of Otago, Dunedin, New Zealand
    ${ }^{3}$ Cooperative Institute for Marine and Atmospheric Research, Pacific Islands Fisheries Science Center, 1845
    Wasp Blvd. Bldg. 176 Honolulu, Hawaii 96818
    ${ }^{4}$ Pacific Community (SPC), Ocean Fisheries Programme (OFP), Noumea, New Caledonia

